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**Nitrogen uptake by the macro-algae *Cladophora coelothrix* and *Cladophora parriaudii*: influence on growth, nitrogen preference and biochemical composition**

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## Abstract

The capacity of macro-algae to remove nutrients means they have the potential to concomitantly bioremediate polluted waters and generate exploitable biomass. The influence of different nitrogen (N) regimes on growth, biochemical composition and bioremediation capacity was studied for two species of the macro-alga *Cladophora*. These were incubated in media containing four single N sources, ammonium ( $\text{NH}_4^+$ ), nitrite ( $\text{NO}_2^-$ ), nitrate ( $\text{NO}_3^-$ ) and urea ( $\text{CO}(\text{NH}_2)_2$ ), each with four nitrogen/phosphorous (N/P) ratios, followed by equimolar dual mixtures of these N sources at two selected N/P ratios. There were clear differences in growth between species, depending upon the nutrient regime. In every instance, the daily growth rate (DGR) of *Cladophora parriaudii* (4.75 – 11.2%) was higher than that of *Cladophora coelothrix* (3.98 – 7.37%) with significance when either  $\text{NO}_2^-$  ( $p = 0.025$ ) or urea ( $p = 0.002$ ) were the employed N form. Differences in algal productivity were reflected in the corresponding N-uptake, whereby *C. parriaudii* consistently removed more N than *C. coelothrix*. There were significant differences in growth ( $p = 0.005$ ) when *C. parriaudii* was cultivated in a single and multi-N source medium:  $\text{NH}_4^+$  was preferentially removed from the medium, whereas urea was typically removed secondarily. However, the presence of urea in the medium enhanced the uptake of the other co-existing N forms and resulted in an increased DGR and yielded a biomass rich in carbohydrates. The relative composition of *C. parriaudii* varied depending upon N/P ratio of the medium, with the final proportion of protein and carbohydrate ranging from 5-15% and 36-54% per unit dry weight, respectively. Results from this study demonstrated that algal strain selection is key to treating waste-streams with specific N profiles. Additionally, the biochemical profile of the biomass produced is dependent on the alga and the N regime, providing the potential for designing processes with specific properties and products.

**Keywords:** *Cladophora*, alga, bioremediation, nitrogen removal, growth rate, biochemical composition

### **Author Contributions:**

M.E.R. is responsible for the integrity of the work as a whole, all named authors contributed materially to the execution of the final manuscript and were involved in the inception, data acquisition, analysis and interpretation, drafting and editing the manuscript.

## 1. Introduction

Nitrogen is typically present in wastewater effluents as either dissolved inorganic nitrogen (DIN), *i.e.* ammonia ( $\text{NH}_3$ ), ammonium ( $\text{NH}_4^+$ ), nitrite ( $\text{NO}_2^-$ ) and nitrate ( $\text{NO}_3^-$ ), or as dissolved organic nitrogen (DON), *i.e.* urea ( $\text{CO}(\text{NH}_2)_2$ ) [1]. Since some of these substances are hazardous and can have an adverse effect on the aquatic environment, the Water Framework Directive (Directive 2000/60/EC) has issued a variety of directives to improve water quality across Europe, which include the Nitrates Directive 1991 and the Dangerous Substances Directive 2006/11/EC. The development and implementation of novel or innovative biological wastewater treatment (WWT) technologies that can remove these pollutants and reduce their potentially deleterious effects is therefore of vital ecological and political importance.

Macro-algae are a diverse collection of macroscopic aquatic organisms the best known of which are the common green, brown and red seaweeds. Some species have high growth rates of 7.1-11.7% per day [2], good nitrogen uptake capacities between 19-96.6% [3] and beneficial utilization of  $\text{CO}_2$  [4]. In addition, the biomass produced by wild harvest or via cultivation could be used as a foodstuff, fertiliser, high-value product, or for conversion into biofuels. These features have stimulated interest in exploring their potential utilisation for bioremediation in biological WWT, *e.g.* in inland and offshore aquaculture systems, including marine integrated multi-trophic aquaculture (IMTA) [5]. Aquaculture is the fastest growing animal food production sector worldwide, therefore improving sustainability through the effective treatment of its waste products is of the utmost importance [5]. In these systems, the production of biomass could concomitantly offer an economic return or negate some of the operational costs of WWT [6]. However, wastewater properties,

including nutrient characteristics, can be highly variable [7], which will influence algal growth rates, biomass production, nutrient uptake and algal composition. Therefore, understanding and optimising these parameters, as well as selecting robust species that maintain good performance, despite potential fluctuations in WWT conditions, are prerequisites for their successful implementation [8].

In an aquaculture WWT scenario, nutrients will be supplied continuously with N concentrations reported in the range of 50-124,900  $\mu\text{M}$  [6, 9]. WWT with algae typically employs outdoor raceway ponds or high-rate algal ponds (HRAPs) with a treatment time of several weeks [10]. However, many studies involving N-uptake by macro-algae are unsuitable for assessing their potential as a tool for nutrient removal, given their experimental design and provision of conditions being unrepresentative of real conditions: these include the supply of low N concentrations ( $<35 \mu\text{M}$ ) and/or the use of short incubation times ( $<24 \text{ h}$ ), with no data provided on algal growth rates [11, 12]. The determination of algal growth is one of the most fundamental parameters in phycological research, used for the comparison between treatments, assessment of biomass production and optimisation of culture conditions [13]. In addition, studies often employ the use of an algal inoculum that has been pre-acclimated in conditions designed to lower their internal nutrient reserves [14]. The subsequent re-introduction to a N replete medium will result in “surge uptake”, where N is removed at a transiently enhanced rate due to the replenishment of internal N pools [15]. This form of uptake may indicate how well suited an individual species is at exploiting pulses of nutrients in an otherwise oligotrophic environment [1], but will be of reduced value in a bioremediation context [10].

Furthermore, the relationship between algal growth and nutrient uptake is very complex and requires further research. Macro-algal nutrient uptake is dependent upon N concentration and typically follows a Michaelis-Menten-type curve, whereby N removal follows a linear trend with increasing external N concentration until a plateau is reached [3, 16]. The rate and maximum amount of N removal achieved is dependent upon biological, physical, and chemical factors, which includes the form of N present [16, 17]. For example, *Gracilaria vermiculophylla* removed 100% of  $\text{NH}_4^+$  in contrast with ~40% of  $\text{NO}_3^-$  [14]. A similar trend was also observed when cultivated in N mixtures, where  $\text{NH}_4^+$  is generally removed ahead of  $\text{NO}_3^-$  [16, 18], or that  $\text{NO}_3^-$  uptake was suppressed when  $\text{NH}_4^+$  was present [19]. Nitrogen source preference by macro-algae is complex and may be determined by: species-specific morphology, shore-line position, nutritional or life-history, the physiological capability and energetic requirement to uptake and assimilate certain nitrogen forms [1, 19, 20].

Fundamentally, mechanisms for N uptake are dependent on the N form present and are geared towards the same objective, namely: an N form enters the cell across the cell membrane, passively or actively, whereupon it is reduced to  $\text{NH}_4^+$  and amino acids, before assimilation into macromolecules, like proteins, for growth [16]. Assimilation of  $\text{NO}_3^-$  occurs by sequentially reducing  $\text{NO}_3^-$  to  $\text{NO}_2^-$ , followed by  $\text{NO}_2^-$  to  $\text{NH}_4^+$ , via the action of N reducing enzymes [10]. Urea is converted into  $\text{NH}_4^+$  by urease [21]. However, there is a dearth of studies involving macro-algal removal of other N forms that are also present in wastewaters, such as  $\text{NO}_2^-$  and urea, as well as mixtures containing these N forms [12, 22]. These N sources are components of the overall dissolved N pool and may play an important role in the algal N budget [12] and also the efficiency of nutrient removal.

In addition to growth rate and nutrient uptake capacity, macro-algal biochemical composition is also affected by nutrient regime. Furthermore, the composition of biomass is important to determine in order to assess their suitability for commercial applications. Several studies have demonstrated seasonal changes in macro-algal chemical composition elicited by differences in environmental conditions as well as nutrient supply [23]. It has been shown experimentally that the tissue N content of macro-algal biomass increases with increasing concentrations of N in their growth medium [24], in contrast with an accumulation of polysaccharides under N-limitation [25]. However, there is still a lack of publically available data on the influence of nutrient regimes in regards to nutrient type, concentration and N/P ratio on macro-algal growth, N uptake, and biochemical composition.

The primary aim of this study was to examine the ability of two robust marine species of algae from the genus *Cladophora* for N removal when subjected to different nutrient regimes and their influence on growth and biochemical composition, with a view for future use in bioremediation. *Cladophora* was selected as a model because of the high growth rates and propensity to bloom in nutrient rich conditions. They are also renowned for their tolerance to a wide range of abiotic factors and resistance to grazers. In addition, *Cladophora* are globally distributed in fresh-, brackish-, and marine-waters: this ubiquity means that local strains or species could be employed, on a case-by-case basis, avoiding the introduction of non-native or alien species [26-28]. These features make *Cladophora* a suitable candidate for investigation into a broad range of applied biotechnological applications, including bioremediation [8]. To explore this potential, cultivation medium was formulated with four different N/P ratios, in order to simulate those found in wastewater streams [29], with nitrogen supplied in both organic and inorganic forms found in wastewaters, specifically  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$  and urea. In addition, N forms were supplied



either singly or dually in equimolar concentrations to elucidate if there was preferential N uptake. This is the first study of its kind to cultivate and characterise algae under the conditions described above. Algal performance was assessed on the basis of growth, nitrogen removal and biochemical composition of the biomass produced.

## 2. Materials and Methods

### 2.1. Macro-algal strains and culture conditions

Algal strains studied were obtained from the Culture Collection of Algal and Protozoa (CCAP), at the Scottish Association for Marine Science (SAMS, UK). These included two marine isolates of the genus *Cladophora*: *Cladophora coelothrix* CCAP 505/10 and *Cladophora parriaudii* CCAP 505/09. Cultures were grown in a nutrient enriched media prepared as follows for all experiments: 33.5 g L<sup>-1</sup> artificial seawater (Instant Ocean, UK), pH adjusted to 7.95 ± 0.08 with buffering provided by the addition of 2 mM Tris-HCl; trace metals (initial concentration = 11 µM Na<sub>2</sub>EDTA, 11 µM FeCl<sub>3</sub>·6H<sub>2</sub>O, 40 nM CuSO<sub>4</sub>·5H<sub>2</sub>O, 76.5 nM ZnSO<sub>4</sub>·7H<sub>2</sub>O, 42 nM CoCl<sub>2</sub>·6H<sub>2</sub>O, 910 nM MnCl<sub>2</sub>·4H<sub>2</sub>O, 25 nM Na<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O); vitamins (initial concentration = 2.97 nM Thiamine HCl, 0.004 nM Cyanocobalamin, 0.02 nM Biotin); phosphorous was added as dihydrogenphosphate dihydrate (NaH<sub>2</sub>PO<sub>4</sub>·2H<sub>2</sub>O) to give an initial P concentration of 80 µM. Nitrogen type and concentration was varied and was supplied as either ammonium chloride, sodium nitrate, sodium nitrite, or urea at molar N/P ratios of 2/1, 12/1, 22/1, and 32/1, equating to initial N concentrations of 160, 960, 1760, and 2560 µM, respectively, with concentrations based upon those found in aquaculture effluents [6, 9]. For the dual nitrogen experiments, N was added in equimolar combinations

at two N/P ratios (*i.e.* 2/1 and 12/1). All chemicals used were obtained from Fisher Scientific, UK.

The cultures were incubated in an illuminated shaker (Sartorius Stedim Biotech, Germany) at 100 rpm at a temperature of 24°C, under an 18/6 h (Light/Dark) photoperiod with 30-40  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  of photosynthetically active radiation (PAR: 400-700) (LM-100 Light Meter, Amprobe, Germany). The algae studied were pre-acclimated in their respective experimental conditions and medium for seven days, after which time 35.7 mg fresh weight (FW) sub-samples were inoculated into triplicate 100 mL flasks containing 50 mL of the experimental medium and then incubated, as outlined above, for 14 days. Additional flasks containing the experimental media only acted as negative controls, whereas those containing biomass and f/2 medium served as a positive control.

## *2.2. Measurement of biomass and its constituents*

Cultures were sampled six times during the study period: days 0, 2, 5, 9, 12, and 14 whereby they were removed aseptically in a laminar flow cabinet (MSC Advantage, Thermo Scientific) for FW determination and water chemistry analysis. On each sampling day, FW was determined by placing the biomass into a reticulated spinner (Chef'N Salad Spinner), spinning for 90 s to remove residual water and weighed gravimetrically in an analytical balance (PS-60, Fisher Brand, UK) [13]. Biomass was then returned to its original, corresponding flask and cultivated under the conditions described above. After the experimental period, the algal cultures were harvested and rinsed for 3-5 seconds with deionised water, from a wash bottle, to remove extracellular salts and nutrients, FW was

determined, the samples frozen at -18°C and then freeze-dried overnight or until a <5% variation in mass was achieved (Modulyo 4K Freeze-Dryer).

Biochemical composition analysis was performed on lyophilised material. Protein was extracted using hot-TCA and an overnight incubation in Lowry Reagent D, and quantified using the Lowry assay, as described by Slocombe *et al.* [30]. Total soluble carbohydrate of the biomass was extracted and quantified using a modified phenol-sulphuric acid method derived from that of Dubois *et al.* [31]. Biomass (5 mg) was hydrolysed with 2.5 mL 1M H<sub>2</sub>SO<sub>4</sub> and autoclaved at 121°C for 15 min (TCR/40/H, Touchclave-R, LTE Scientific, UK). Hydrolysates were cooled to room temp (RT) and centrifuged for 2 min at 15,600 g (5414 Microcentrifuge, Eppendorf®). The assay was performed on 30 µL of supernatant with the direct addition of 0.5 mL of 4% phenol (w/v), followed by 2.5 mL of conc. H<sub>2</sub>SO<sub>4</sub> (> 96%). The reaction was cooled to room temperature and read at 490 nm (Helios Gamma UV-vis spectrophotometer, Thermo Scientific), which was calibrated against a 0-5 mg L<sup>-1</sup> glucose calibration curve. Blanks were composed of deionised water in place of the hydrolysate.

### 2.3. Water chemistry analysis

For each sampling day analysis of nutrients present in solution was performed. Soluble NO<sub>2</sub><sup>-</sup> and NO<sub>3</sub><sup>-</sup> were measured using Ion Chromatography (883 Basic IC Plus, Metrohm, UK) [13]. Total Ammonia Nitrogen (TAN) and phosphate (PO<sub>4</sub><sup>3-</sup>) were determined colourimetrically using proprietary test-kits (1.00683.0001 and 1.14848.0001, Spectroquant®) and following the manufacturer's instructions.

Urea was determined in water samples using a method derived from that of Mulvenna and Savidge [32]. Urea reagents were freshly prepared on each day of analysis and were: Urea Reagent A (1.7 g of diacetylmonoxime in 46 mL dH<sub>2</sub>O, made up to 50 ml with thiosemicarbazide solution: 0.475 g in 100 mL dH<sub>2</sub>O), and Urea Reagent B (31.26 mL dH<sub>2</sub>O was acidified with 40 mL conc. H<sub>2</sub>SO<sub>4</sub>, once cooled 66 µL of ferric (III) chloride solution is added: 0.15 g in 10 mL dH<sub>2</sub>O). Samples were diluted with deionised water to fall within the calibration range, and 2 mL of sample was added to a reaction tube. To this, 143 µL of Urea Reagent A was added and immediately mixed (Vortex 3, IKA®), followed by 458 µL of Urea Reagent B and mixed again. Teflon-lined screw-caps were loosely added and the samples were incubated for 30 mins at 85°C (Gallenkamp Plus II Incubator, Sanyo, UK). Samples were quickly cooled in a water bath for a total of 10 min, replacing the water after 5 minutes. The samples were then read in a spectrophotometer at 520 nm (Helios Gamma UV-vis spectrophotometer, Thermo Scientific) and compared against a 0-5 mg L<sup>-1</sup> urea calibration curve. Blanks were composed of deionised water in place of the sample.

## 2.4. Data analysis

Algal growth is expressed as daily growth rate (DGR) and was determined using the equation (1) as recommended by Yong *et al.* [33]:

$$DGR = \left[ \left( \frac{W_t}{W_0} \right)^{1/d} - 1 \right] \times 100 \quad (1)$$

Where  $W_t$  and  $W_0$  are the final and initial FW mass, and  $d$  is the time in days.

Protein and carbohydrate content of the biomass is portrayed as % DW. Data on water chemistry were converted to molar N form and expressed either as total N uptake ( $\mu\text{M}$ ), or as a proportion of N removed from the growth medium (%), both relative to values obtained from corresponding negative controls. All experiments were performed in triplicate and the experimental error was calculated and expressed as one standard deviation (SD). Within a single N/P ratio, comparison of DGR, nutrient uptake, and biochemical composition was performed using a 2-sample *t*-test. A one-way ANOVA with Tukey's post hoc analysis was used to determine differences in DGR of *C. parriaudii* between multiple N sources provided at the same concentration. Relationships between DGR and N removal, and protein to carbohydrate content were described using Pearson's correlation, *r*. Levels of significance were set at  $p < 0.05$ ; all statistical analyses were performed using Minitab® Statistical Software version 17.

### 3. Results and discussion

In this study, two species of *Cladophora* were investigated for N removal, growth rate, and biochemical composition after 14 days incubation under different nutrient regimes that were designed to simulate a broad range of wastewaters. The only experimental variable was nitrogen addition, with growth media formulated with four different N/P ratios and four different sources of nitrogen:  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$ , or urea. This study was performed in two main sections. Firstly, N sources were added singly for comparison between species on a fundamental level. Secondly, media was formulated with dual nitrogen sources in equimolar concentrations, provided at a 2/1 and 12/1 N/P ratio, to elucidate the N source preference of *C. parriaudii*. The removal of N was attributed to the algal strains studied by

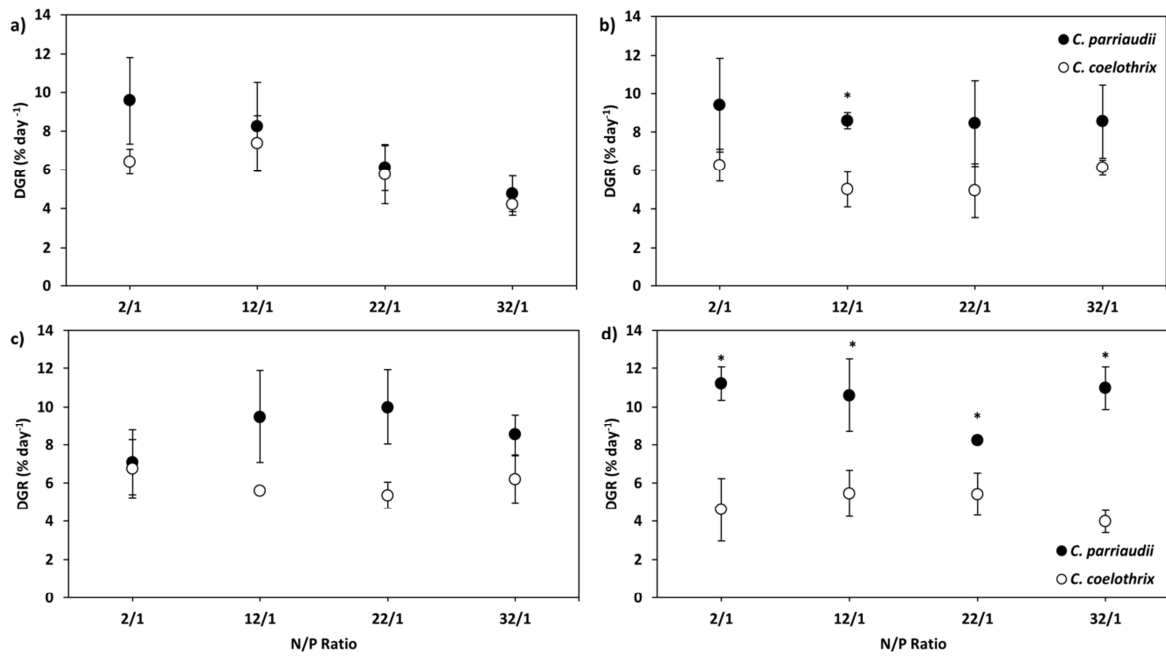
comparison against negative control flasks where nutrient removal was not observed (see Supplementary Data Fig. S1). In addition, several samples, across different nutrient regimes and sampling days, were analysed for the presence of other nitrogen forms: in no instance was any other form of N present than that which was originally added (data not shown). Positive control cultures grew consistently throughout the experimental time-frame, indicating that no unexpected changes in growth occurred (Supplementary Data Fig. S2). In addition,  $\text{PO}_4^{3-}$  removal was never greater than 53%, therefore, neither species of *Cladophora* would have suffered from P-limitation (Supplementary Data Fig. S3 and S4). The addition of Tris-HCl buffered the cultivation media, which prevented excessive shifts in pH caused by algal growth, with final pH values (*i.e.* measured on day 14) of  $8.18 \pm 0.08$  and  $8.4 \pm 0.25$  measured for *C. coelothrix* and *C. parriaudii*, respectively.

### 3.1. Uptake of single nitrogen sources

Experiments were performed in order to determine whether two closely related species of *Cladophora* are able to sequester different N sources indiscriminately, across a range of concentrations, and if this influences growth. Single sources of N were added at four different N/P ratios, with the DGR and N uptake determined (Fig. 1 and 2).

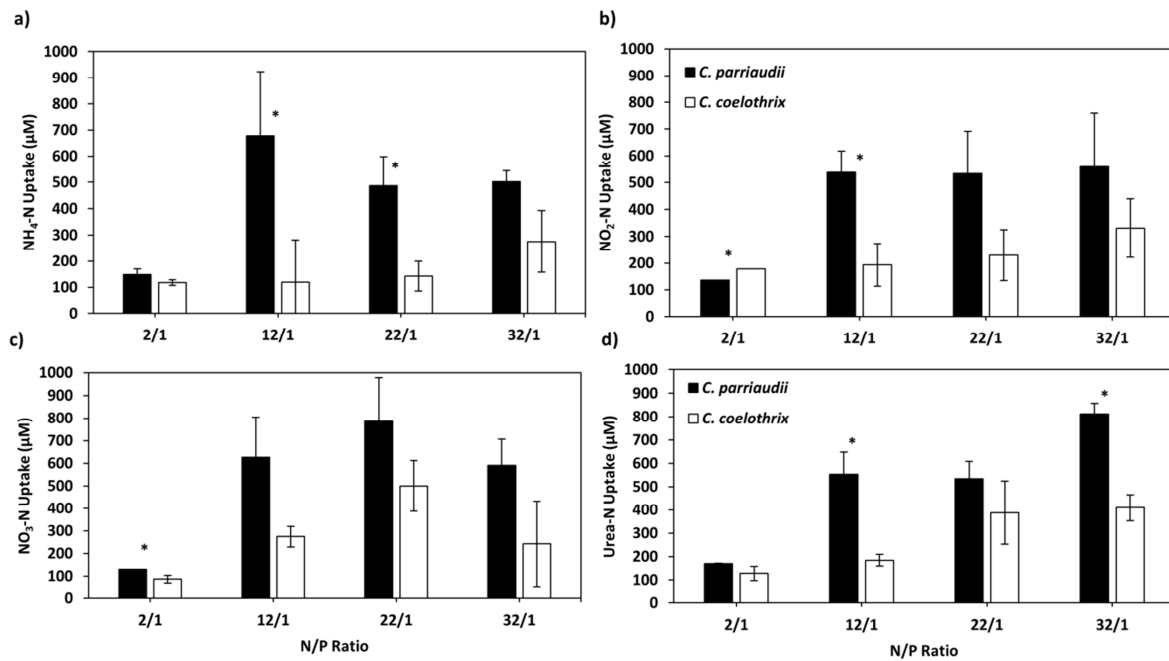
Both species studied exhibited growth and N uptake irrespective of nutrient regime. *C. parriaudii* had a higher DGR (Fig. 1) and N-uptake (Fig. 2) than *C. coelothrix* under all nutrient regimes tested. Differences in DGR were statistically significant at 12/1  $\text{NO}_2^-$  ( $p = 0.025$ ) and for all urea concentrations tested ( $p = 0.002 - 0.049$ ) (Fig. 1). A reduction in growth rate for both species with increasing concentrations of  $\text{NH}_4^+$  was observed (Fig. 1a). This was most likely due to the increasing concentration of the toxic un-ionised  $\text{NH}_3$  form

that exists in equilibrium with  $\text{NH}_4^+$  [34]. Previous studies have reported toxic effects of  $\text{NH}_3$  with algal cultures at concentrations ranging from 2.34 - 2000  $\mu\text{M}$  [35, 36], and  $\text{NH}_3$  concentrations in this study fell within this range, varying between 0-250  $\mu\text{M}$ . Disparities in growth between the two species were generally reflected in the amount of N removed (Fig. 2). For example, at a 12/1 ratio with  $\text{NO}_2^-$ , DGR for *C. parriaudii* was 8.6% as opposed to 5% for *C. coelothrix*, with corresponding  $\text{NO}_2\text{-N}$  uptake of 538 ( $\pm 79.9$ )  $\mu\text{M}$  and 193 ( $\pm 80.3$ )  $\mu\text{M}$ , respectively (Fig. 1b and 2b). Overall, differences in DGR and N uptake observed may be accounted for by interspecific variations in physiology, capacity to utilise N forms, growth strategy, as well as morphology [20, 37]. For instance, Luo *et al.* [37] reported that *Ulva prolifera* had higher rates of growth and nutrient uptake than *Ulva linza* across a range of temperatures, irradiances, and N concentrations, most likely as a result of them having a different thallus morphology. A similar explanation may be valid in this case: Ross *et al.* [13] noted that *C. parriaudii* grows rapidly outwards in a loose skein, whereas *C. coelothrix* grows slowly in tightly knit clusters. The denser growth strategy exhibited by *C. coelothrix*, as opposed to *C. parriaudii*, will likely enhance the effects of growth-limiting factors, such as self-shading and low nutrient availability within the tightly knit colony, as observed in other members of Cladophoraceae [38, 39].



**Fig. 1.** The DGR of *C. parriaudii* (black) and *C. coelothrix* (white) cultivated for 14 days in different media formulations with N/P ratios of 2/1, 12/1, 22/1, and 32/1, equivalent to initial N concentrations of 160, 960, 1760, and 2560  $\mu\text{M}$ , respectively. N was supplied as  $\text{NH}_4^+$  (a),  $\text{NO}_2^-$  (b),  $\text{NO}_3^-$  (c), or urea (d) ( $n = 3$ , error bars = 1 SD). Statistically different DGR between species are indicated \* ( $p < 0.05$ ).





**Fig. 2.** The total N uptake by *C. parriaudii* (black) and *C. coelothrix* (white) when cultivated for 14 days in different media formulations with N/P ratios of 2/1, 12/1, 22/1, and 32/1, equivalent to initial N concentrations of 160, 960, 1760, and 2560  $\mu\text{M}$ , respectively. Nitrogen was supplied as  $\text{NH}_4^+$  (a),  $\text{NO}_2^-$  (b),  $\text{NO}_3^-$  (c), or urea (d) ( $n = 3$ , error bars = 1 SD). Statistically different total N uptake between species are indicated \* ( $p < 0.05$ ).

Greater differences in performance between the two species occurred when cultivated on urea. Both the highest and lowest DGR was achieved with 11.2% by *C. parriaudii* and 3.98% by *C. coelothrix*, respectively. This indicates that two closely related species of *Cladophora* have different capacities for N source utilisation. Literature on macro-algal urea removal is sparse in comparison to  $\text{NO}_3^-$  or  $\text{NH}_4^+$  [1]. However, Phillips and Hurd [1] observed negative values for urea uptake by inter-tidal macro-algal species during winter conditions, but noted uptake saturation during summer months with increasing urea concentrations. The authors hypothesized that the energetically expensive enzyme urease, required to convert urea to

$\text{NH}_4^+$ , may be down-regulated during winter months when abiotic conditions are sub-optimal for growth. A similar concept may apply here, where there was a strongly positive correlation between the DGR and N uptake by *C. parriaudii* at each nutrient ratio, with Pearson's  $r$  values of 0.645-0.848 and  $p = <0.001 - 0.024$ . However, no such relationship existed with *C. coelothrix*,  $p = 0.656 - 0.999$ , which suggests that the growth rate of *C. parriaudii* was strongly influenced by the nutrient regime, whereas the growth of *C. coelothrix* is likely to be more strongly influenced by another parameter, such as light intensity. This suggests that *C. coelothrix* may have under-expressed urease production as a consequence of sub-optimal cultivation conditions and the low rate of urea assimilation that occurred may have been for maintenance or housekeeping purposes.

A common trend in N uptake was not shared between *C. coelothrix* and *C. parriaudii* (Fig. 2), most likely due to the physical and biological factors mentioned above. In the case of *C. parriaudii*, N uptake followed a Michaelis-Menten-type trend, generally plateauing at a ratio of 12/1 and beyond, which is comparable to observations reported by other studies [3, 16]. The kinetics of nutrient uptake were described in detail by Harrison and Hurd [17], defining this plateau as a period of "internally controlled uptake", with the activity of N reducing enzymes limiting the rate of further N uptake.

Due to the lower rates of growth and N uptake, *C. coelothrix* was not investigated further. However, *C. coelothrix* has been reported to be a robust species [13] and merits further study in conditions more representative of WWT, where it may exhibit a higher degree of tolerance and growth in comparison to *C. parriaudii* [40]. In addition, as N removal by *C. parriaudii* plateaued beyond a 12/1 ratio, only ratios of 2/1 and 12/1 were investigated for the remainder of this study.

### 3.2. Uptake of nitrogen combinations

Combinations of different nitrogen sources were added to the growth matrices to determine whether *C. parriaudii* is capable of complimentary N uptake and what effect this has on DGR (Fig. 3) and total N uptake (Fig. 4). Data regarding N removal at the 2/1 ratio are available as Supplementary Data (Fig. S5): these are not shown, as the trends are similar to the 12/1 ratio, albeit with complete N removal achieved by day 9.

Generally, cultures that have removed the most N also tend to have a higher DGR. The N source appeared to have a greater effect on the DGR of *C. parriaudii* than N/P ratio (Fig. 3): no significant differences in growth were observed between N/P ratios, whilst there was significance in the DGR values within a given N source. For example, the DGR of *C. parriaudii* when cultivated under a 2/1 ratio with urea as the common N source (Fig. 3d) ranged from 7.8-11.2% ( $p = 0.005$ ).

The inclusion of urea in the media tended to enhance both the growth and N uptake by *C. parriaudii* and may be explained when considering its influence on biochemical composition of the biomass, which will be discussed later. Greater total N removal was achieved by *C. parriaudii* when incubated in  $\text{NH}_4$ -Urea (79.8%) as opposed to  $\text{NH}_4^+$  alone (67.7%) and mixtures of  $\text{NH}_4$ - $\text{NO}_2$  (32.7%) and  $\text{NH}_4$ - $\text{NO}_3$  (50.4%) (Fig. 4a). The same trend was observed in media combinations with  $\text{NO}_2^-$  or  $\text{NO}_3^-$  as the common N source (Fig. 4b and c). Interestingly, growth and N uptake was greater in urea mixtures in comparison to a single addition of urea: the DGR of single urea at 12/1 was 9.2%; however, this increased to 10.6%, 10.6%, and 11.2% with combinations of  $\text{NH}_4$ -Urea,  $\text{NO}_2$ -Urea, and  $\text{NO}_3$ -Urea, respectively (Fig. 3d). As growth and N removal are strongly related, a corresponding trend was found in urea uptake. The lowest total N removal was observed with a single urea addition (61.4%), as opposed to

combinations containing urea:  $\text{NH}_4$ -Urea with 79.8%,  $\text{NO}_2$ -Urea with 77.5%, and  $\text{NO}_3$ -Urea with 65.2% (Fig. 4d). Overall, these results indicate that *C. parriaudii* is capable of complimentary N uptake and that this can enhance growth. In a previous study, Bracken and Stachowicz [20] reported that of eight local species tested, neither *Cladophora* nor *Prionitis* exhibited the greatest N removal in monoculture. However, *in situ*, they were in fact the most dominant in the Bodega Marine Reserve, California, partly attributed to their ability to utilise multiple nitrogen forms when available.

In contrast with urea, incorporating either  $\text{NH}_4^+$  or  $\text{NO}_2^-$  into a mixture generally resulted in reduced DGR: for example, a treatment of 12/1  $\text{NO}_2$ - $\text{NO}_3$  resulted in one of the lowest observed rates of growth (6.9%) and total N removal (47.7%). Both  $\text{NO}_2^-$  and  $\text{NH}_3$  (which co-exists in equilibria with  $\text{NH}_4^+$ ) are toxic to algae [34, 41], and possibly impaired the functionality of the organism.

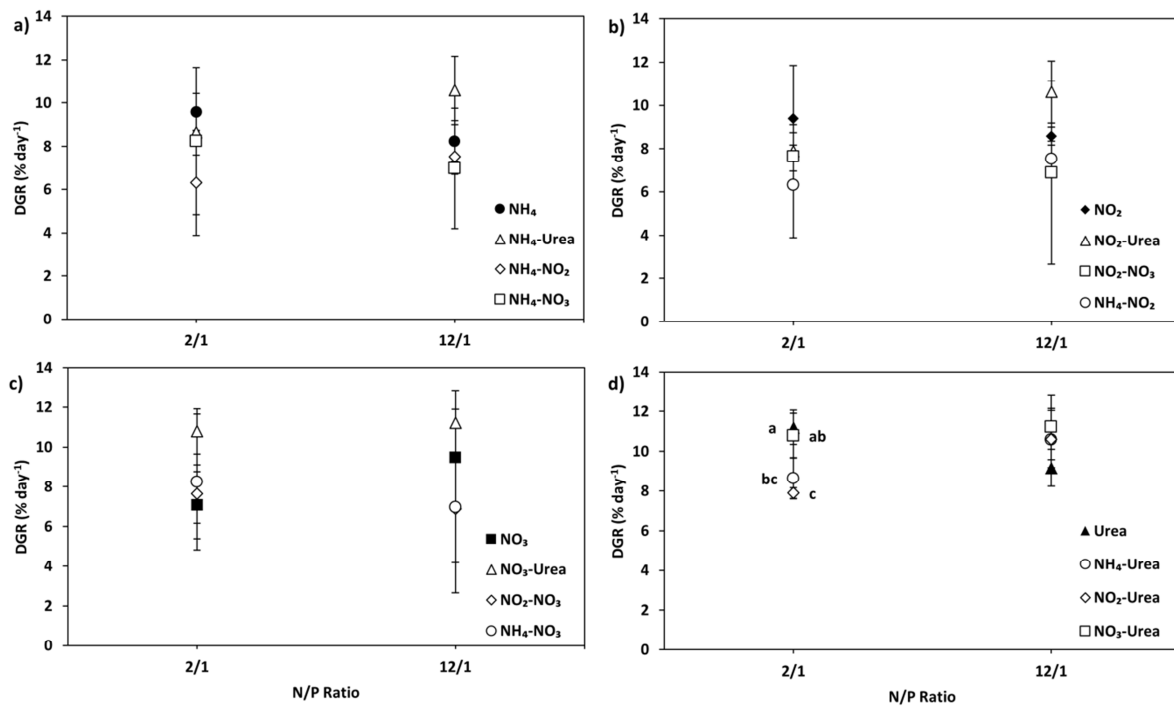
Differences in algal performance may be a result of an inherent preference for a certain nitrogen form, which can be elucidated by measuring temporal N removal of the component N sources.

### 3.3. Preferential nitrogen uptake

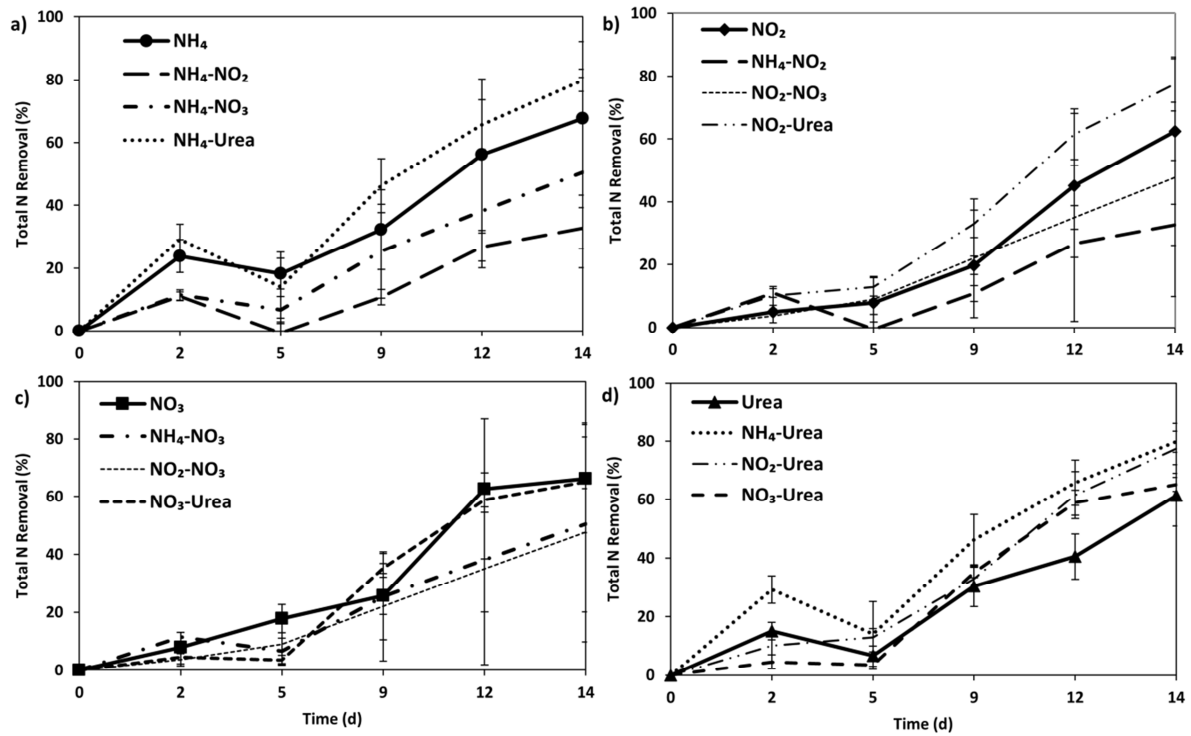
To determine whether *C. parriaudii* exhibited any preference for a particular N source, the removal of the component N sources from the dual N media was investigated (Fig. 5). The trends between 2/1 and 12/1 N removal preference were very similar; however, as a lesser concentration of N was added in the 2/1 regime, the relationships were achieved more

rapidly making them more difficult to discern. For this reason only 12/1 have been included (Fig. 5), and the 2/1 data is available as Supplementary Data (Fig. S6).

There are clear differences in N uptake preference for *C. parriaudii*:  $\text{NH}_4^+$  was removed at a greater rate and to a higher degree than any other N source it was mixed with (Fig. 5a, b, and c). A preference for  $\text{NH}_4^+$  over  $\text{NO}_3^-$  has been reported elsewhere for both eukaryotic algae and cyanobacteria [14, 42, 43]. However, there are very few studies on macro-algal preference in dual N media formulations involving alternative nitrogen sources, such as urea and  $\text{NO}_2^-$  [12, 22]. This preference for  $\text{NH}_4^+$  is presumed to be because it is less energetically expensive to assimilate than the other N sources, as  $\text{NO}_2^-$  or  $\text{NO}_3^-$  are more oxidised forms of nitrogen that need to be reduced to  $\text{NH}_4^+$ , whereas urea requires different enzymes for its assimilation [14, 21, 44]. Additionally, algal-bacterial interactions may also play a role, such as nutrient mineralization [45].



**Fig. 3.** The DGR of *C. parriaudii* cultivated for 14 days in different media formulations with 2/1 and 12/1 N/P ratios, equivalent to initial N concentrations of 160 and 960  $\mu\text{M}$ , respectively. N was added as  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$ , or urea, either singularly or dually in equimolar concentrations. The figures have been grouped by common N source as  $\text{NH}_4^+$  (a),  $\text{NO}_2^-$  (b),  $\text{NO}_3^-$  (c), and urea (d), with the darkened shape denoting the common type ( $n = 3$ , error bars = 1 SD). Treatments which do not share a letter are significantly different, whereas, treatments that share a letter or have no letter are not significantly different ( $p < 0.05$ ).

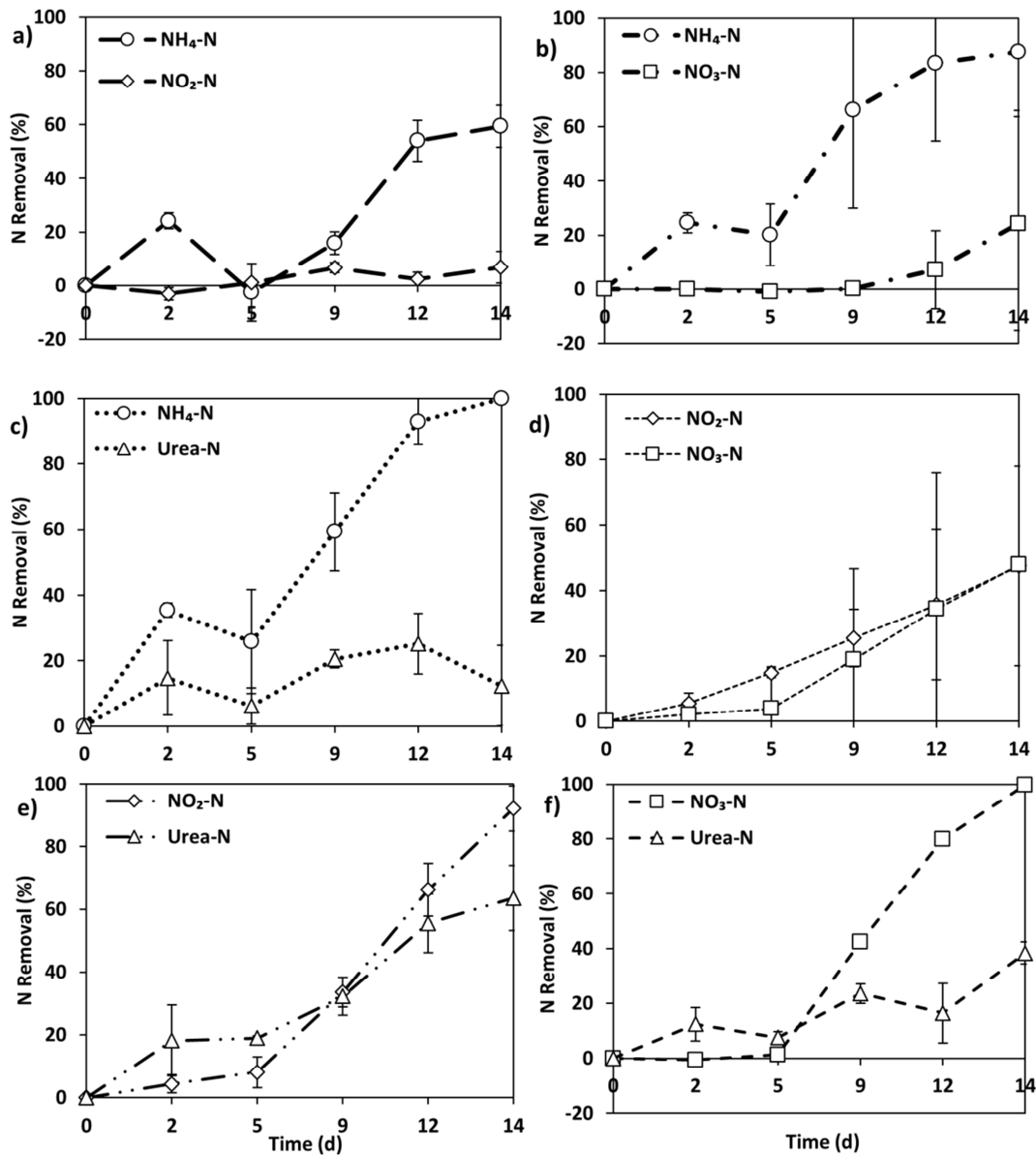


**Fig. 4.** The temporal total N removal by *C. parriaudii* cultivated for 14 days in different media formulations with a 12/1 N/P ratio, equivalent to an initial N concentration of 960  $\mu\text{M}$ . Nitrogen was added as  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$ , or urea, either singularly or dually in equimolar concentrations. The figures have been grouped by common N source as  $\text{NH}_4^+$  (a),  $\text{NO}_2^-$  (b),  $\text{NO}_3^-$  (c), and urea (d) with the darkened shape and solid line denoting the common type for each corresponding graph ( $n = 3$ , error bars = 1 SD).

Similarities between the removal of  $\text{NO}_2^-$  and  $\text{NO}_3^-$  by *C. parriaudii* cultivated in  $\text{NO}_2^-\text{-NO}_3^-$  media were observed, with an overall equal removal of 47% for each, despite a slight initial delay in  $\text{NO}_3^-$  uptake (Fig. 5d). Furthermore, their removal when in conjunction with other forms, in this case with  $\text{NH}_4^+$  and urea, were similar. There was an initial 5-day lag period for  $\text{NO}_2^-$  and  $\text{NO}_3^-$  uptake before 92.2% and 99.8% were removed, respectively, when urea was the additional nitrogen type (Fig. 5e and f). However, only 6.8% and 24.3% of  $\text{NO}_2^-$  and  $\text{NO}_3^-$ ,

respectively, were removed when  $\text{NH}_4^+$  was also present in the media (Fig. 5a and b). Raven *et al.* [44] suggested that  $\text{NO}_3^-$  uptake could be completely suppressed when in the presence of 1000-2000  $\mu\text{M}$   $\text{NH}_4^+$ , with the most plausible reason being that  $\text{NH}_4^+$  inhibits the biosynthesis of nitrate/nitrite reductase or urease enzymes [21]. In this study,  $\text{NO}_3^-$  removal only occurred after the concentration of  $\text{NH}_4^+$  had reduced to 95-207  $\mu\text{M}$ , typically achieved between days 5 and 9. However, in the  $\text{NH}_4\text{-NO}_2$  treatment, the concentration of  $\text{NH}_4^+$  never went below 154  $\mu\text{M}$  and translated into a low removal of  $\text{NO}_2^-$  of <7%. This suggests that there is a threshold  $\text{NH}_4^+$  concentration above which  $\text{NO}_2^-$  and  $\text{NO}_3^-$  removal is suppressed. This was further highlighted at the 2/1 ratio (Fig. S6) where the onset of  $\text{NO}_2^-$  and  $\text{NO}_3^-$  removal from the  $\text{NH}_4\text{-NO}_2$  and  $\text{NH}_4\text{-NO}_3$  mixtures occurred once  $\text{NH}_4^+$  was almost depleted. The presence of  $\text{NH}_4^+$  in the media was considered to be the causative agent of the poor removal of  $\text{NO}_2^-$  and  $\text{NO}_3^-$ . These virtually identical trends in removal of  $\text{NO}_2^-$  and  $\text{NO}_3^-$  were assumed to be due to their chemical similarity and their common metabolic pathway for nitrogen reduction and assimilation.





**Fig. 5.** The preferential removal of component N sources by *C. parriaudii* cultivated for 14 days in different media formulations with an N/P ratio of 12/1, equivalent to an initial N concentration of 960  $\mu$ M. Nitrogen was added as NH<sub>4</sub><sup>+</sup> (circle), NO<sub>2</sub><sup>-</sup> (diamond), NO<sub>3</sub><sup>-</sup> (square), or urea (triangle) in equimolar concentrations. The graphs have been grouped by their dual nitrogen additions as NH<sub>4</sub>-NO<sub>2</sub> (a), NH<sub>4</sub>-NO<sub>3</sub> (b), NH<sub>4</sub>-Urea (c), NO<sub>2</sub>-NO<sub>3</sub> (d), NO<sub>2</sub>-Urea (e), and NO<sub>3</sub>-Urea (f) ( $n = 3$ , error bars = 1 SD).

A >92% removal of  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ , and  $\text{NO}_3^-$  was observed, when added in an equimolar combination with urea; however, the corresponding removal of urea was comparatively low, amounting to 12.4%, 63.6%, and 38.5%, respectively (Fig. 5c, e, and f). This was a somewhat surprising result given the relatively high rate of urea removal when added as a single source (61.4%) (Fig. 4d), and the correspondingly high DGRs when it was incorporated into dual N media (Fig. 3d). Tyler *et al.* [12] observed enhanced uptake of urea in the presence of  $\text{NH}_4^+$  by *Gracilaria vermiculophylla* and *Ulva lactuca*, suggesting that each N form has a distinct uptake mechanism allowing their simultaneous removal. This was in contrast with results observed in this study where urea uptake was suppressed in the presence of other N sources, especially  $\text{NH}_4^+$ . The discrepancy between these findings is likely due to differences in experimental design and cultivation conditions: Tyler *et al.* [12] employed a shorter time-frame and a much greater biomass/nutrient ratio, with an inoculum that was previously maintained in low nutrient seawater, meaning that surge uptake would have played a prominent role in nutrient removal.

### 3.4. Biochemical composition

It should be noted that, despite being commonly used for biochemical analysis, the colourimetric methods used for biochemical determination in this work have their limitations, for example the non-determination of uronic acids from the carbohydrate assay [23, 30, 31, 46].

A strong inverse correlation between protein and carbohydrate content ( $r = -0.585$ ,  $p < 0.001$ ) was observed: generally, algal cultures maintained in a 2/1 ratio had a low protein and high carbohydrate content, whereas at a 12/1 ratio, they had a high protein and

reduced carbohydrate composition (Table 1). At a 2/1 ratio, N was generally completely removed after 9 days (Fig. S5), likely resulting in N-deprivation and hence a lower protein content, whereas at the 12/1 ratio N was never depleted and therefore cultures would not have been N-limited and protein synthesis will have continued (Fig. 4). The influence of nutrient supply on biochemical composition has been observed elsewhere, where macroalgae have been found to accumulate storage polysaccharides under N-deprivation, with a shift towards the synthesis of proteins and pigments when N is sufficient [25, 47]. In this study, the highest protein content at both ratios was found in *C. parriaudii* maintained under  $\text{NH}_4\text{-NO}_2$ , with values of 10.8% and 15% at 2/1 and 12/1, respectively, coincident with the lowest DGR and total N uptake (Fig. 3a and 4a). Meanwhile, the highest carbohydrate content was 54%, achieved in flasks where the alga was cultivated under a 2/1 ratio with  $\text{NO}_2\text{-Urea}$ , where complete N removal was attained after 9 days.

It appears that N uptake, as seen between different N/P ratios, has a greater influence on biochemical composition than N form (Table 1), despite Corey *et al.* [18] finding that the tissue N content of the biomass of *Palmaria palmata* significantly changed from 3.1% to 4.1% when cultivated under  $\text{NO}_3^-$  or  $\text{NH}_4^+$ , respectively. In their study, there was however a significantly greater removal of  $\text{NH}_4^+$  in comparison to  $\text{NO}_3^-$  [18], therefore, it seems more likely that an algal species preference for an N form will influence the uptake rate and absolute N removal, which will in turn affect the biomass composition, in contrast with the N form having a direct influence on biochemical composition.

The inclusion of urea in dual N media combinations resulted in carbohydrate rich biomass. For example, three of the four highest yields at a 2/1 ratio (48.4-54%) and the three highest at a 12/1 ratio (40.6-42.8%) were obtained when urea was part of the growth medium

(Table 1). Considering that urea is an organic molecule its influence upon the algal biochemical composition may be slightly more complex. Both Choo *et al.* [48] and Raven *et al.* [49] have described multiple mechanisms for inorganic carbon acquisition by *Cladophora* sp., therefore the capacity to utilise carbon from urea is a distinct possibility. The conversion of urea to  $\text{NH}_4^+$ , via enzymatic activity, results in the formation of 2  $\text{NH}_3$  and one  $\text{CO}_2$  molecule [21]. This  $\text{CO}_2$  formation may enhance photosynthetic activity and facilitate carbohydrate construction [50]. This theory of complimentary carbon and N sequestration may also explain the enhanced DGR and N uptake by *C. parriaudii* when cultivated in growth medium containing urea (Fig. 3d and 4d).

**Table 1.** The protein and carbohydrate content per unit DW of *C. parriaudii* cultivated for 14 days in different media formulations with 2/1 and 12/1 N/P ratios, equivalent to initial N concentrations of 160 and 960  $\mu\text{M}$ , respectively. Nitrogen was added as  $\text{NH}_4^+$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$ , or urea, singly or dually in equimolar concentrations ( $n = 3$ ,  $\pm 1$  SD). Values highlighted \* are significantly different from one another within a nitrogen source ( $p < 0.05$ ).

Nitrogen Source	Protein (% DW)		Carbohydrate (% DW)	
	2/1	12/1	2/1	12/1
$\text{NH}_4^+$	6.2 ( $\pm 1.8$ )*	13 ( $\pm 0.7$ )*	51.1 ( $\pm 2$ )*	31.1 ( $\pm 1.3$ )*
$\text{NO}_2^-$	5.4 ( $\pm 1.3$ )*	10.9 ( $\pm 1.2$ )*	41.7 ( $\pm 6.7$ )	38.5 ( $\pm 3.1$ )
$\text{NO}_3^-$	6.8 ( $\pm 1.2$ )	9.7 ( $\pm 0.5$ )	41.1 ( $\pm 2$ )*	32 ( $\pm 1.9$ )*
Urea	6.5 ( $\pm 0.6$ )	8 ( $\pm 0.9$ )	43.8 ( $\pm 11.9$ )	37.6 ( $\pm 3.6$ )
$\text{NH}_4\text{-NO}_2$	10.8 ( $\pm 3.3$ )	15 ( $\pm 0.4$ )	45.6 ( $\pm 6.2$ )	39.2 ( $\pm 1.5$ )
$\text{NH}_4\text{-NO}_3$	8.3 ( $\pm 2.9$ )	12.1 ( $\pm 1.9$ )	47.2 ( $\pm 5.9$ )	36.1 ( $\pm 6.5$ )
$\text{NH}_4\text{-Urea}$	5.1 ( $\pm 0.2$ )*	11.4 ( $\pm 1.8$ )*	51.5 ( $\pm 0.9$ )	40.6 ( $\pm 4.5$ )
$\text{NO}_2\text{-NO}_3$	8.4 ( $\pm 3$ )	11.9 ( $\pm 1.1$ )	38.8 (5.6)	38.2 ( $\pm 2.2$ )
$\text{NO}_2\text{-Urea}$	6.5 ( $\pm 1.1$ )*	12 ( $\pm 0.9$ )*	54 ( $\pm 8.1$ )	42.7 ( $\pm 2.2$ )
$\text{NO}_3\text{-Urea}$	6.1 ( $\pm 0.2$ )*	10.7 ( $\pm 0.4$ )*	48.4 ( $\pm 1.3$ )	42.8 ( $\pm 3.1$ )

## 4. Conclusions

There were several key conclusions from this study. Firstly, there were species-specific differences in growth, N-uptake, and ability to utilise certain N forms between two species of *Cladophora*. Furthermore, *C. parriaudii* is capable of complimentary N-uptake and its growth is strongly influenced by N type, rather than concentration, showing a preference towards  $\text{NH}_4^+$ , potentially due to its reduced form and lower energetic requirement for its assimilation, whereas urea was removed secondarily. However, the presence of urea in the medium enhanced the uptake of other N forms present, possibly due to complimentary carbon sequestration. Whereas, the total amount and rate of uptake of both  $\text{NO}_2^-$  and  $\text{NO}_3^-$  were almost identical. Finally, different nutrient regimes elicited a change in the final composition of *C. parriaudii* biomass, with N concentration having a greater influence than N form. This demonstrates that a sea change in algal cultivation practices is required and should include selecting species for biological WWT purposes that are robust, flexible, and can simultaneously utilise multiple carbon and nitrogen forms. Future work on a larger-scale and in conditions more representative of WWT are required and in a broader algal biotechnology context, alternative, cheaper, and more easily assimilable N sources could be employed. Nutrient regimes could be tailored to reduce costs and produce biomass with required commercial attributes.

## References

- [1] J.C. Phillips, C.L. Hurd, Kinetics of nitrate, ammonium, and urea uptake by four intertidal seaweeds from New Zealand, *J. Phycol.* 40 (2004) 534-545.
- [2] Y.-F. Yang, X.-G. Fei, J.-M. Song, H.-Y. Hu, G.-C. Wang, I.K. Chung, Growth of *Gracilaria lemaneiformis* under different cultivation conditions and its effects on nutrient removal in Chinese coastal waters, *Aquaculture*. 254 (2006) 248-255.
- [3] M. Jimenez del Rio, Z. Ramazanov, G. Garcia-Reina, *Ulva rigida* (Ulvales, Chlorophyta) tank culture as biofilters for dissolved inorganic nitrogen from fishpond effluents, *Fifteenth International Seaweed Symposium*, Springer, 1996, pp. 61-66.
- [4] A.D. Hughes, K.D. Black, I. Campbell, K. Davidson, M.S. Kelly, M.S. Stanley, Does seaweed offer a solution for bioenergy with biological carbon capture and storage?, *Greenhouse Gases: Science and Technology*. 2 (2012) 402-407.
- [5] J. Bostock, B. McAndrew, R. Richards, K. Jauncey, T. Telfer, K. Lorenzen, D. Little, L. Ross, N. Handisyde, I. Gatward, *Aquaculture: global status and trends*, *Philos. T. Roy. Soc. B*. 365 (2010) 2897-2912.
- [6] L. Hayashi, N.S. Yokoya, S. Ostini, R.T. Pereira, E.S. Braga, E.C. Oliveira, Nutrients removed by *Kappaphycus alvarezii* (Rhodophyta, Solieriaceae) in integrated cultivation with fishes in re-circulating water, *Aquaculture*. 277 (2008) 185-191.
- [7] D. Dominguez, W. Gujer, Evolution of a wastewater treatment plant challenges traditional design concepts, *Water Res.* 40 (2006) 1389-1396.
- [8] P.H. de Paula Silva, R. De Nys, N.A. Paul, Seasonal growth dynamics and resilience of the green tide alga *Cladophora coelothrix* in high-nutrient tropical aquaculture, *Aquaculture Environment Interactions*. 2 (2012) 253-266.
- [9] A.E. Ghaly, M. Kamal, N.S. Mahmoud, Phytoremediation of aquaculture wastewater for water recycling and production of fish feed, *Environ. Int.* 31 (2005) 1-13.
- [10] T. Cai, S.Y. Park, Y. Li, Nutrient recovery from wastewater streams by microalgae: status and prospects, *Renew. Sust. Energ. Rev.* 19 (2013) 360-369.
- [11] D.T. Dy, H.T. Yap, Surge ammonium uptake of the cultured seaweed, *Kappaphycus alvarezii* (Doty) Doty (Rhodophyta: Gigartinales), *J. Exp. Mar. Biol. Ecol.* 265 (2001) 89-100.
- [12] A.C. Tyler, K.J. McGlathery, S.A. Macko, Uptake of urea and amino acids by the macroalgae *Ulva lactuca* (Chlorophyta) and *Gracilaria vermiculophylla* (Rhodophyta), *Mar. Ecol-Prog. Ser.* 294 (2005) 161-172.
- [13] M.E. Ross, M.S. Stanley, J.G. Day, A.J.C. Semião, A comparison of methods for the non-destructive fresh weight determination of filamentous algae for growth rate analysis and dry weight estimation, *J. App. Phycol.* (2017) 1-12.
- [14] M.H. Abreu, R. Pereira, A. Buschmann, I. Sousa-Pinto, C. Yarish, Nitrogen uptake responses of *Gracilaria vermiculophylla* (Ohmi) Papenfuss under combined and single addition of nitrate and ammonium, *J. Exp. Mar. Biol. Ecol.* 407 (2011) 190-199.
- [15] M.F. Pedersen, J. Borum, Nutrient control of estuarine macroalgae: growth strategy and the balance between nitrogen requirements and uptake, *Mar. Ecol-Prog. Ser.* 161 (1997) 155-163.
- [16] C. Wang, A. Lei, K. Zhou, Z. Hu, W. Hao, J. Yang, Growth and nitrogen uptake characteristics reveal outbreak mechanism of the opportunistic macroalga *Gracilaria tenuistipitata*, *PLoS One*. 9 (2014) e108980.
- [17] P.J. Harrison, C.L. Hurd, Nutrient physiology of seaweeds: application of concepts to aquaculture, *Cah. Biol. Mar.* 42 (2001) 71-82.

- [18] P. Corey, J.K. Kim, J. Duston, D.J. Garbary, B. Prithiviraj, Bioremediation potential of *Palmaria palmata* and *Chondrus crispus* (Basin Head): effect of nitrate and ammonium ratio as nitrogen source on nutrient removal, *J. Appl. Phycol.* 25 (2013) 1349-1358.
- [19] A. Smit, Nitrogen uptake by *Gracilaria gracilis* (Rhodophyta): adaptations to a temporally variable nitrogen environment, *Bot. Mar.* 45 (2002) 196-209.
- [20] M.E. Bracken, J.J. Stachowicz, Seaweed diversity enhances nitrogen uptake via complementary use of nitrate and ammonium, *Ecology*. 87 (2006) 2397-2403.
- [21] P. Syrett, J. Leftley, Nitrate and urea assimilation by algae, in: N. Sunderland, E. Simon, J. Heslop-Harrison, P. Brian, D. Boulter (Eds.) *Perspectives in experimental biology*, Pergamon Press, Oxford, UK, 2016, pp. 221-234.
- [22] N.A. Waser, K. Yin, Z. Yu, K. Tada, P.J. Harrison, D.H. Turpin, S.E. Calvert, Nitrogen isotope fractionation during nitrate, ammonium and urea uptake by marine diatoms and coccolithophores under various conditions of N availability, *Mar. Ecol-Prog. Ser.* 169 (1998) 29-41.
- [23] P. Schiener, K.D. Black, M.S. Stanley, D.H. Green, The seasonal variation in the chemical composition of the kelp species *Laminaria digitata*, *Laminaria hyperborea*, *Saccharina latissima* and *Alaria esculenta*, *J. App. Phycol.* 27 (2015) 363-373.
- [24] S. Sode, A. Bruhn, T.J. Balsby, M.M. Larsen, A. Gotfredsen, M.B. Rasmussen, Bioremediation of reject water from anaerobically digested waste water sludge with macroalgae (*Ulva lactuca*, Chlorophyta), *Bioresource Technol.* 146 (2013) 426-435.
- [25] A.J. Smit, B.L. Robertson, D.R. du Preez, Influence of ammonium-N pulse concentrations and frequency, tank condition and nitrogen starvation on growth rate and biochemical composition of *Gracilaria gracilis*, *J. App. Phycol.* 8 (1997) 473-481.
- [26] K.-S. Choo, P. Snoeijs, M. Pedersén, Oxidative stress tolerance in the filamentous green algae *Cladophora glomerata* and *Enteromorpha ahlneri*, *J. Exp. Mar. Biol. Ecol.* 298 (2004) 111-123.
- [27] S.B. Zulkifly, J.M. Graham, E.B. Young, R.J. Mayer, M.J. Piotrowski, I. Smith, L.E. Graham, The genus *Cladophora* Kützting (Ulvophyceae) as a globally distributed ecological engineer, *J. Phycol.* 49 (2013) 1-17.
- [28] D.M. Wotton, C. O'Brien, M.D. Stuart, D.J. Fergus, Eradication success down under: heat treatment of a sunken trawler to kill the invasive seaweed *Undaria pinnatifida*, *Mar. Pollut. Bull.* 49 (2004) 844-849.
- [29] M.S. Islam, Nitrogen and phosphorus budget in coastal and marine cage aquaculture and impacts of effluent loading on ecosystem: review and analysis towards model development, *Mar. Pollut. Bull.* 50 (2005) 48-61.
- [30] S.P. Slocombe, M. Ross, N. Thomas, S. McNeill, M.S. Stanley, A rapid and general method for measurement of protein in micro-algal biomass, *Bioresource Technol.* 129 (2013) 51-57.
- [31] M. Dubois, K.A. Gilles, J.K. Hamilton, P. Rebers, F. Smith, Colorimetric method for determination of sugars and related substances, *Anal. Chem.* 28 (1956) 350-356.
- [32] P.F. Mulvenna, G. Savidge, A modified manual method for the determination of urea in seawater using diacetylmonoxime reagent, *Estuar. Coast. Shelf S.* 34 (1992) 429-438.
- [33] Y.S. Yong, W.T.L. Yong, A. Anton, Analysis of formulae for determination of seaweed growth rate, *J. App. Phycol.* 25 (2013) 1831-1834.
- [34] A. König, H.W. Pearson, S.A. Silva, Ammonia toxicity to algal growth in waste stabilization ponds, *Water Sci. Technol.* 19 (1987) 115-122.
- [35] A. Abeliovich, Y. Azov, Toxicity of ammonia to algae in sewage oxidation ponds, *Appl. Environ. Microb.* 31 (1976) 801-806.
- [36] T. Källqvist, A. Svenson, Assessment of ammonia toxicity in tests with the microalga, *Nephroselmis pyriformis*, Chlorophyta, *Water Res.* 37 (2003) 477-484.
- [37] M.B. Luo, F. Liu, Z.L. Xu, Growth and nutrient uptake capacity of two co-occurring species, *Ulva prolifera* and *Ulva linza*, *Aquat. Bot.* 100 (2012) 18-24.

- [38] D. Krause-Jensen, K. McGlathery, S. Rysgaard, P.B. Christensen, Production within dense mats of the filamentous macroalga *Chaetomorpha linum* in relation to light and nutrient availability, *Mar. Ecol-Prog. Ser.* 134 (1996) 207-216.
- [39] I. Wallentinus, Comparisons of nutrient uptake rates for Baltic macroalgae with different thallus morphologies, *Mar. Biol.* 80 (1984) 215-225.
- [40] P.H. de Paula Silva, S. McBride, R. de Nys, N.A. Paul, Integrating filamentous 'green tide' algae into tropical pond-based aquaculture, *Aquaculture*. 284 (2008) 74-80.
- [41] S. Philips, H.J. Laanbroek, W. Verstraete, Origin, causes and effects of increased nitrite concentrations in aquatic environments, *Reviews in Environmental Science and Biotechnology*. 1 (2002) 115-141.
- [42] X. Fan, D. Xu, Y. Wang, X. Zhang, S. Cao, S. Mou, N. Ye, The effect of nutrient concentrations, nutrient ratios and temperature on photosynthesis and nutrient uptake by *Ulva prolifera*: implications for the explosion in green tides, *J. App. Phycol.* 26 (2014) 537-544.
- [43] M. Ohmori, K. Ohmori, H. Strotmann, Inhibition of nitrate uptake by ammonia in a blue-green alga, *Anabaena cylindrica*, *Arch. Microbiol.* 114 (1977) 225-229.
- [44] J.A. Raven, B. Wollenweber, L.L. Handley, A comparison of ammonium and nitrate as nitrogen sources for photolithotrophs, *New Phytol.* 121 (1992) 19-32.
- [45] J.J. Cole, Interactions between bacteria and algae in aquatic ecosystems, *Annu. Rev. Ecol. Syst.* 13 (1982) 291-314.
- [46] A.R. Angell, L. Mata, R. de Nys, N.A. Paul, The protein content of seaweeds: a universal nitrogen-to-protein conversion factor of five, *J. Appl. Phycol.* 28 (2016) 511-524.
- [47] T. Chopin, T. Gallant, I. Davison, Phosphorus and nitrogen nutrition in *Chondrus crispus* (Rhodophyta): effects on total phosphorus and nitrogen content, carrageenan production, and photosynthetic pigments and metabolism, *J. Phycol.* 31 (1995) 283-293.
- [48] K.-S. Choo, P. Snoeijs, M. Pedersén, Uptake of inorganic carbon by *Cladophora glomerata* (Chlorophyta) from the Baltic Sea, *J. Phycol.* 38 (2002) 493-502.
- [49] J. Raven, J. Beardall, H. Griffiths, Inorganic C-sources for *Lemanea*, *Cladophora* and *Ranunculus* in a fast-flowing stream: measurements of gas exchange and of carbon isotope ratio and their ecological implications, *Oecologia*. 53 (1982) 68-78.
- [50] T. Berman, S. Chava, Algal growth on organic compounds as nitrogen sources, *J. Plankton Res.* 21 (1999) 1423-1437.

## Web References

[http://ec.europa.eu/environment/water/water-nitrates/index\\_en.html](http://ec.europa.eu/environment/water/water-nitrates/index_en.html) (last accessed, 26/06/2017)

[http://ec.europa.eu/environment/water/water-dangersub/lib\\_dang\\_substances.htm](http://ec.europa.eu/environment/water/water-dangersub/lib_dang_substances.htm) (last accessed, 26/06/2017)